

## A REVIEW OF METHOD DEVELOPMENT AND METHOD VALIDATION OF FLUCONAZOLE BY RP-HPLC METHOD

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**ABSTRACT:** For the simultaneous detection of fluconazole in bulk and pharmaceutical dose forms, a simple, quick, precise, accurate, and sensitive reverse phase liquid chromatographic approach has been established. Waters ODS C18 RP column, 250 mm 4.6 mm, particle size 5m i.e., UV detection at 274 nm, and a flow rate of 1.0 ml/min were used to standardize the chromatographic procedure. Both bulk and pharmaceutical dose forms of fluconazole were satisfactorily quantified using the suggested approach. Fluconazole was reported to have a linearity range of 6- 16% g/ml. It was determined that the correlation coefficient was 0.995. For fluconazole, 2.90 g/ml was determined to be the LOQ. The lowest detectable concentration of fluconazole was 0.90 g/ml. Research into deterioration was also created. Guidelines from the International Council for Harmonization (ICH) were also used to calculate precision, accuracy, robustness, and other analytical metrics.

**KEYWORDS:** Fluconazole, HPLC, linearity, accuracy, precision, degradation studies.

**INTRODUCTION:** As a synthetic triazole derivative antifungal drug, fluconazole has been demonstrated to be effective against a broad variety of systemic and superficial fungal infections after oral and intravenous treatments. Its chemical name is 2-(2,4-difluorophenyl)-1,3-bis(1H-1,2,4-triazole-1-yl)-2-propanol. Some of the techniques used to analyze fluconazole include gas chromatography (GC) and high performance liquid chromatography (HPLC) for the evaluation of fluconazole in biological fluids; HPLC for eye drops and creams; UV spectrophotometry for syrups, capsules, and intravenous solution; and microbiological test for capsules.

The weight of one mole of fluconazole is 306.271 grams. C<sub>13</sub>H<sub>12</sub>F<sub>2</sub>N<sub>6</sub>O is the chemical formula for this compound. The chemical makeup of fluconazole is

Figure No.1: structure of fluconazole.

The fluconazole is affected by the reduction in renal function. Therefore, the dose of fluconazole needs to be reduced in patients with impaired renal function. If creatinine clearance is  $\leq 50$  ml/min, the clinician must reduce the dose to 50%. The plasma half-life of fluconazole is approximately 30 hours.

## EXPERIMENTAL

### Apparatus

The liquid chromatography equipped with water OSD column C<sub>18</sub> 250mm × 4.66mm and the particle size is 5 µm.

### Reagents and materials

The drugs used were of AR grade. The tablets used is diflucon 150 mg

S.NO.	Name	Specifications		Manufacturer/Supplier
		Purity	Grade	
1.	Doubled distilled water	99.9%	HPLC	Sd fine-Chem ltd; Mumbai
2.	HPLC Grade Water	99.9%	HPLC	Sd fine-Chem ltd; Mumbai
3.	Methanol	99.9%	HPLC	Loba Chem; Mumbai.
4.	Hydrochloric Acid	99.9	A.R.	Sd fine-Chem ltd; Mumbai
5.	Acetonitrile	99.9%	HPLC	Loba Chem; Mumbai.
6.	Sodium Hydroxide	99.9	A.R.	Sd fine-Chem ltd; Mumbai

**Table No.1: list of chemicals used**

### SOLUBILITY

Solubility studies

SOLVENT	SOLUBILITY
Ethanol	Soluble
DMSO	Soluble
Dimethyl formamide	Soluble
Aqueous Buffers	Sparingly soluble
Methanol	Soluble
Water	Slightly Soluble
Ethyl Acetate	Soluble

**Table No.2: list of solvents**

**Chromatographic conditions**

The chromatographic conditions were optimized by different means.  
(Using different column, different mobile phase, different flow rate,  
different detection wavelength & different diluents for sample  
preparation etc.

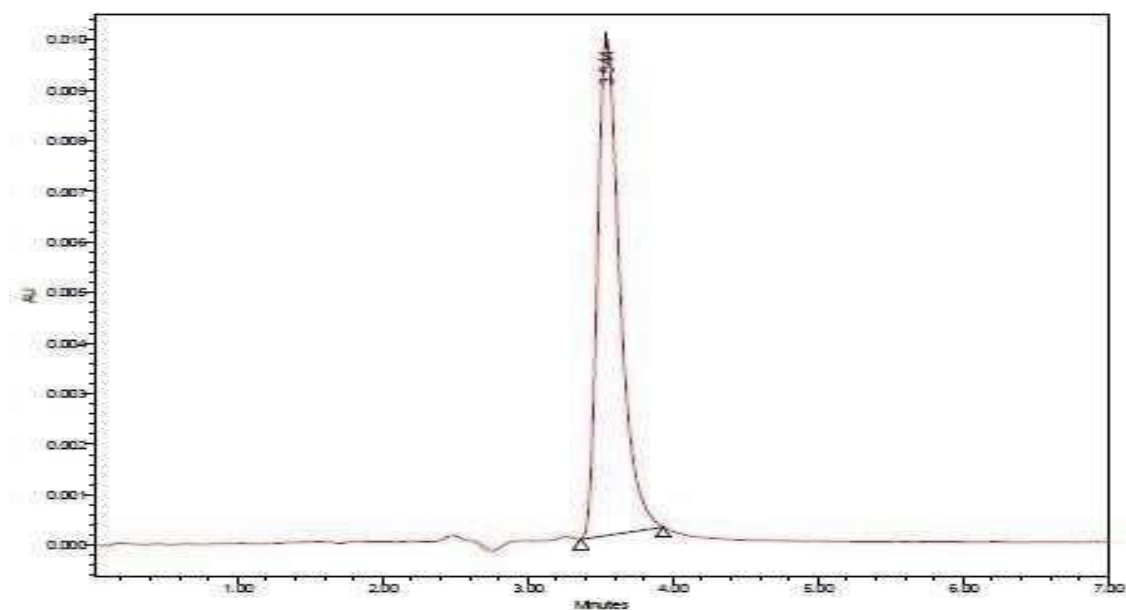
<b>Column Used</b>	<b>Mobile Phase</b>	<b>Flow Rate</b>	<b>Wave length</b>	<b>Observation</b>	<b>Result</b>
Symmetry ODS RP  C <sub>18</sub> ,5m, 15mm x 4.6mm i.d.	Water :ACN  = 50 : 50	0.8 ml/min	274nm	Low response	Method rejected

Symmetry ODS RP C <sub>18</sub> ,5 m, 15mm x 4.6mm i.d.	Methanol : Water = 60 : 40	1.0 ml/min	274nm	Very low response	Method rejected
Symmetry ODS RP C <sub>18</sub> ,5 m, 15mm x 4.6mm i.d.	ACN: Methanol = 70 : 30	1.0 ml/min	274nm	Tailing peak	Method rejected
Symmetry ODS RP C <sub>18</sub> ,5 m, 15mm x 4.6mm i.d.	ACN: Phosphate buffer = 45:55	1.0 ml/ min	274nm	Broad Peak	Method rejected
Symmetry ODS RP C <sub>18</sub> ,5 m, 15mm x 4.6mm i.d.	ACN : Phosphate buffer (pH=3.2) =35:65	1.0 ml/ min	274nm	Tailing peak	Method rejected
Symmetry ODS RP C <sub>18</sub> ,5 m, 15mm x 4.6mm i.d.	ACN : Phosphate buffer (pH=3.2) =20:80	1.0 ml/min	274nm	Good Peak	Method accepted

**Table No.3: summary of process optimization**

Drug Name	Rt	Peak Area	Tailing Factor	Theoretical Plates
Fluconazole	3.544	2812469	1.01	1733

**Table No.4: peak results of optimized conditions**



**Fig No.2: chromatogram of fluconazole in optimized conditions**

## **METHOD DEVELOPMENT**

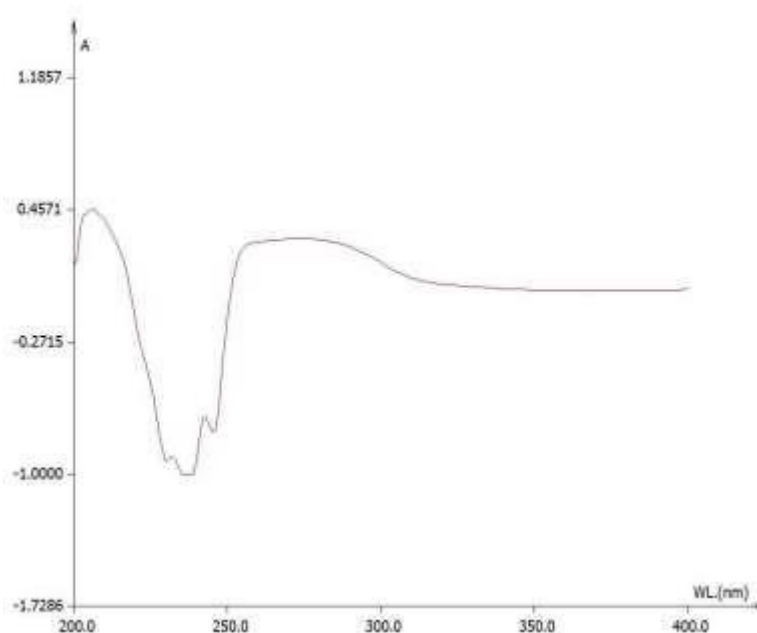
### **Selection of wavelength**

The standard & sample stock solutions were prepared separately by dissolving standard & sample in a solvent in mobile phase diluting with the same solvent. (After optimization of all conditions ) for UV spectrum in the range of 200 to 400nm. This has been performed to know the maxima of Fluconazole, so that the same wave number can be utilized in HPLC UV detector for estimating the Olmesartan. The scanned UV spectrum is attached in the following page

### **Sample & standard preparation for the UV-Spectrophotometer analysis**

10g of fluconazole standard was transferred into 10 ml volumetric flask, dissolved & make up to volume with mobile phase.

Further dilution was done by transferring 1 ml of the above solution into a 10ml volumetric flask and make up to volume with mobile phase.



**Fig No.3: UV spectrum for fluconazole**

Observation: while scanning the fluconazole solution we observed the maxima at 274nm. The UV spectrum has been recorded on T60-LAB INDIA make UV-VIS spectrophotometer model UV-2450.

### **Preparation of mobile phase**

800ml of phosphate buffer and 200 ml of HPLC grade water were mixed well and degassed in ultrasonic water bath for 15 minutes. The solutions was filtered through 0.45 $\mu$ m filter under vacuum filtration.

Final result & discussion: the selected and optimized mobile phase ACN : phosphate buffer (p.H=3.2) = 20:80 and conditions optimized were flow rate (1.0 ml/minute), wavelength (274nm), run time was 7 minutes. Here the peaks were separated and showed better resolution, theoretical plate count and symmetry. The proposed chromatographic conditions were found appropriate for the quantitative determination of the drug.

### Estimation of fluconazole in tablet dosage form

Twenty tablets were taken and the I.P. method was followed to determine the average weight. Above weighed tablets were finally powdered and triturated well. A quantity of powder equivalent to 25 mg of drugs were transferred to 25ml volumetric flask, make and solutions was sonicated for 15 minutes, there after volume was made up to 25 ml with solvent. Then 10 ml of the above solution was diluted to 100 ml with mobile phase. The solution was filtered through a membrane filter (0.45 $\mu$  m) sonicated to degas. The solution prepared was injected in five replicates into the HPLC system and the observations were recorded.

Brand name of Tablets	Labeled amount of Drug (mg)	Mean(SD) amount (mg) found by the proposed method (n=6)	Assay % (SD)
Flucanazole	150	149.98 ( 0.498)	99.9( 0.343)

**Table No.5: recovery data for estimation fluconazole, diflucon 150 mg of fluconazole**

### ACCURACY

To determine the accuracy of the proposed method, recovery studies were carried out by pure drug of fluconazole taken and added to pre-analysed formulation of concentration 10 $\mu$ g/ml. From that percentage recovery values were calculated.

Sample ID	Concentration ( g/ml)		Peak Area	% Recovery of Pure drug	Statistical Analysis
	Amount Added	Amount Found			
S <sub>1</sub> : 80 %	8	8.069	485317	100.862	Mean= 100.5413% S.D. = 0.947606 % R.S.D.= 0.942503
S <sub>2</sub> : 80 %	8	7.958	478751	99.475	
S <sub>3</sub> : 80 %	8	8.103	487312	101.287	

S <sub>4</sub> : 100 %	10	10.048	601947	100.48	Mean= 100.2367% S.D. = 0.650103
S <sub>5</sub> : 100 %	10	10.073	603395	100.73	
S <sub>6</sub> : 100 %	10	9.950	596176	99.50	% R.S.D.= 0.648568
S <sub>7</sub> : 120 %	12	11.985	716127	99.875	Mean= 100.3607% S.D. = 0.555257 % R.S.D. = 0.553262
S <sub>8</sub> : 120 %	12	12.116	723840	100.966	
S <sub>9</sub> : 120 %	12	12.029	718706	100.241	

**Table No.6: Accuracy readings**

### **PRECISION**

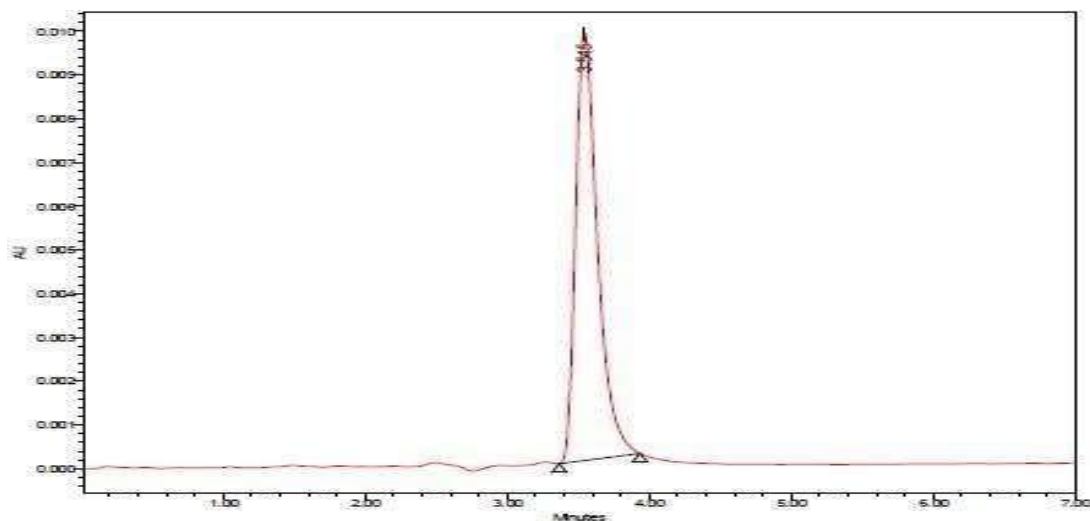
The precision of each method was ascertained separately from the peak areas & retention times obtained by actual determination of six replicates of a fixed amount of drug. Fluconazole (API). The percent relative standard deviation was calculated for fluconazole are presented in below table.

<b>HPLC Injection Replicates of Fluconazole</b>	<b>Retention Time</b>	<b>Peak Area</b>
Replicate – 1	3.545	661022
Replicate – 2	3.537	683137
Replicate – 3	3.543	671941
Replicate – 4	3.538	682245



Replicate – 5	3.542	671941
Replicate – 6	3.550	692444
Average	3.5425	677121.7
Standard Deviation	0.004764	11046.13

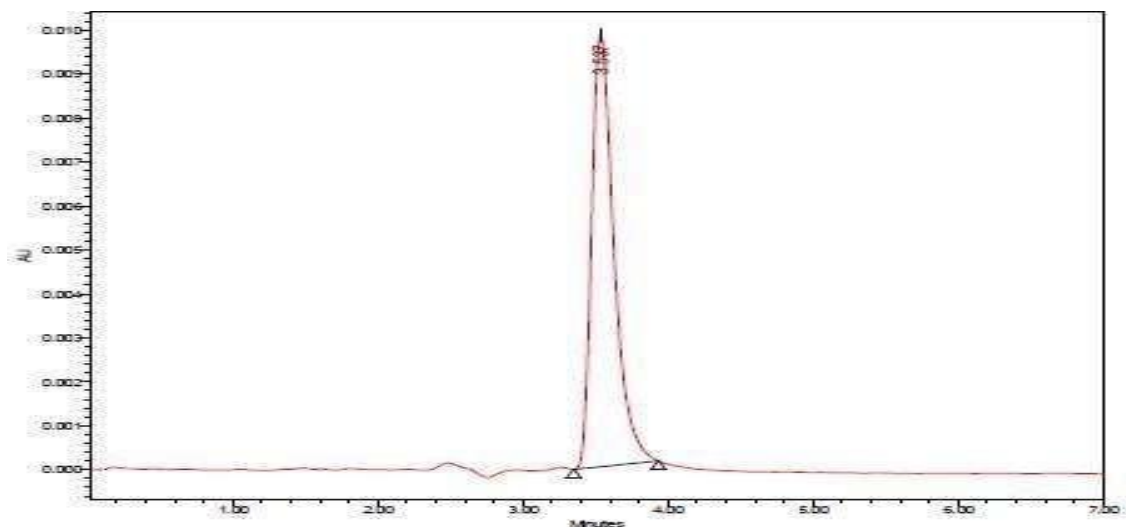
**Table No.7: repeatability readings.**



**Fig-4: Chromatogram of Repeatability-1**

**Table-24: Results of Repeatability-1**

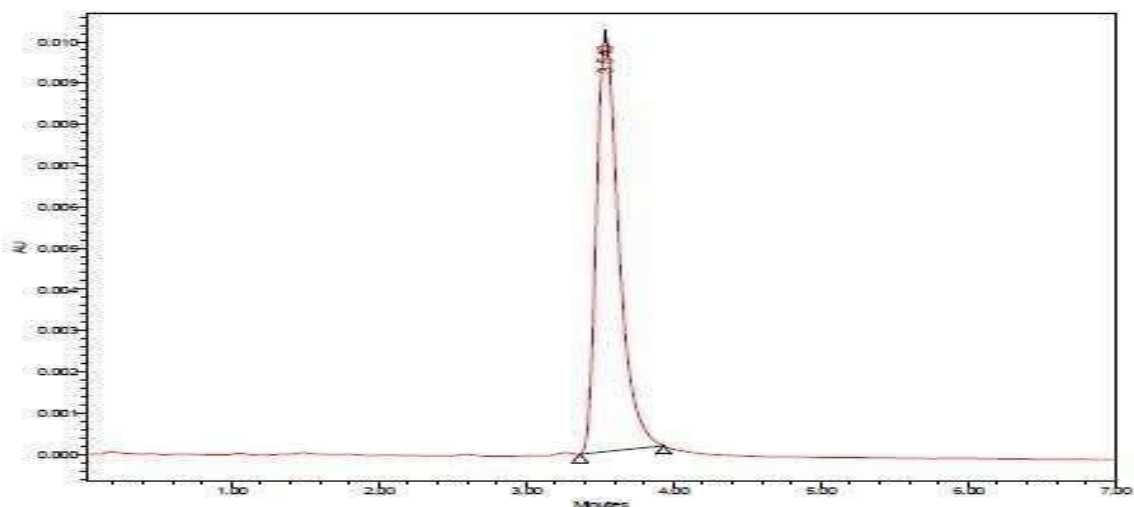
S.No.	Rt	Peak Area	Theoretical Plates	Tailing factor
1	3.545	661022	3169	1.09



**Fig-5: Chromatogram of Repeatability-2**

**Table-25: Results of Repeatability-2**

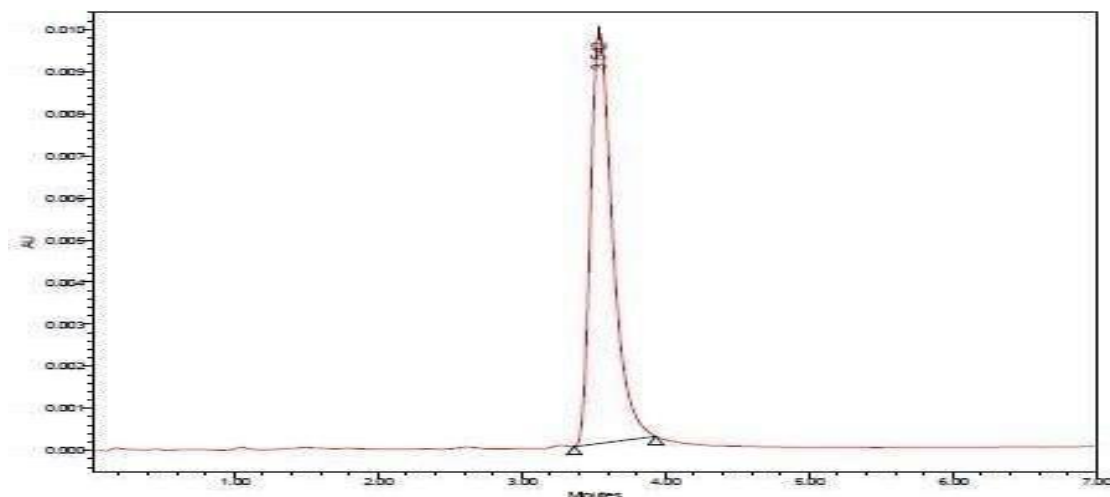
<b>b</b>	<b>Rt</b>	<b>Peak Area</b>	<b>Theoretical Plates</b>	<b>Tailing factor</b>
1	3.537	683137	5214	1.02



**Fig-6: Chromatogram of Repeatability-3**

**Table-26: Results of Repeatability-3**

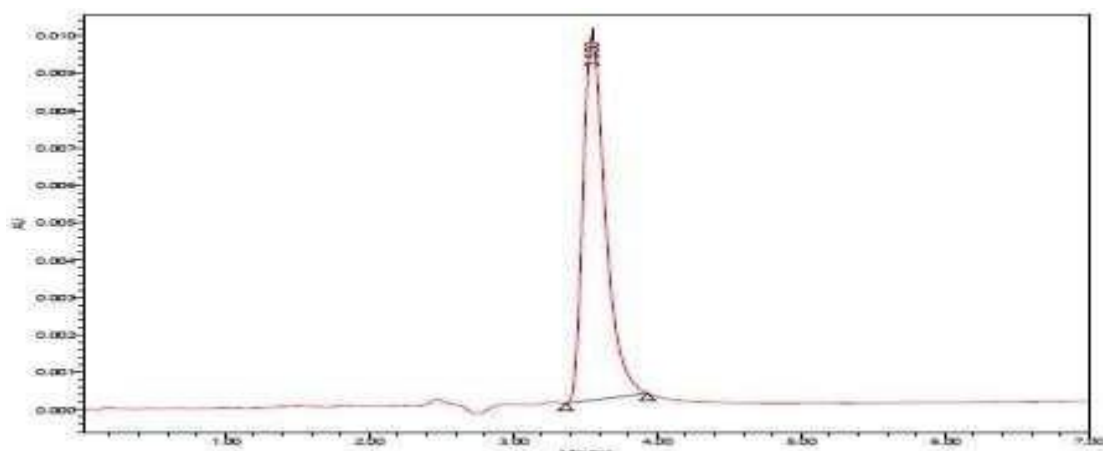
S.No.	Rt	Peak Area	Theoretical Plates	Tailing factor
1	3.543	671941	3368	1.13



**Fig-7: Chromatogram of Repeatability-5**

**Table-27: Results of Repeatability-5**

S.No.	Rt	Peak Area	Theoretical Plates	Tailing factor
1	3.542	671941	6254	0.99



**Fig-8: Chromatogram of Repeatability-6**

**Table-28: Results of Repeatability-6**

S.No.	Rt	Peak Area	Theoretical Plates	Tailing factor
1	3.550	692444	6524	1.25

### Intermediate precision

#### Intra assay & inter assay

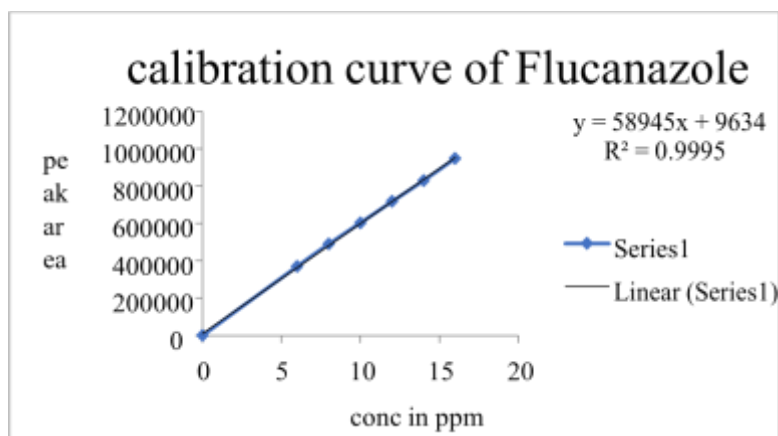
The intra & inter day variation of the method was carried out & the high values of mean assay & low values of standard deviation & % RSD (%RSD < 2%) within a day & day to day variations for fluconazole revealed that the proposed method is precise

Conc. Of Flucanazole (API) (µg/ml)	Observed Conc. Of Flucanazole (µg/ml) by the proposed method			
	Intra-Day		Inter-Day	
	Mean (n=6)	% RSD	Mean (n=6)	% RSD
8	7.76	0.82	8.28	0.98
10	10.16	0.42	9.59	0.23
12	11.68	0.13	12.19	0.33

**Table No.8: results of intra assay & inter assay**

### LINEARITY AND RANGE

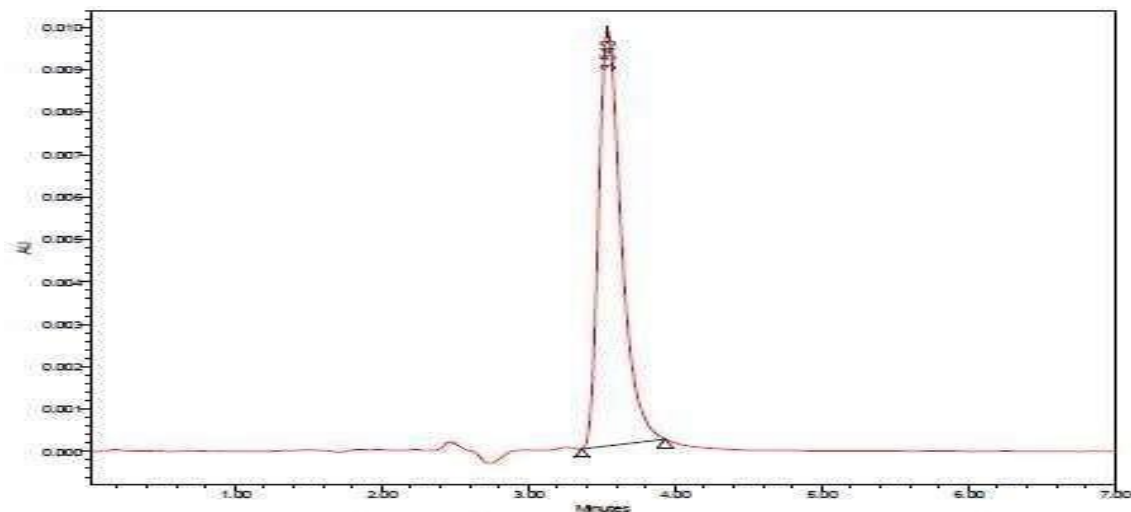
The calibration curve showed the range of 0-16µg/ml, for fluconazole (API) with coefficient of 0.999. a typical calibration curve has the regression equation of  $y = 58954x + 9634$  for



**Fig No.10: calibration curve for fluconazole**

CONC.(µg/ml)	MEAN AUC (n=6)
0	0
6	370200
8	490231
10	602707
12	717538
14	829248
16	947852

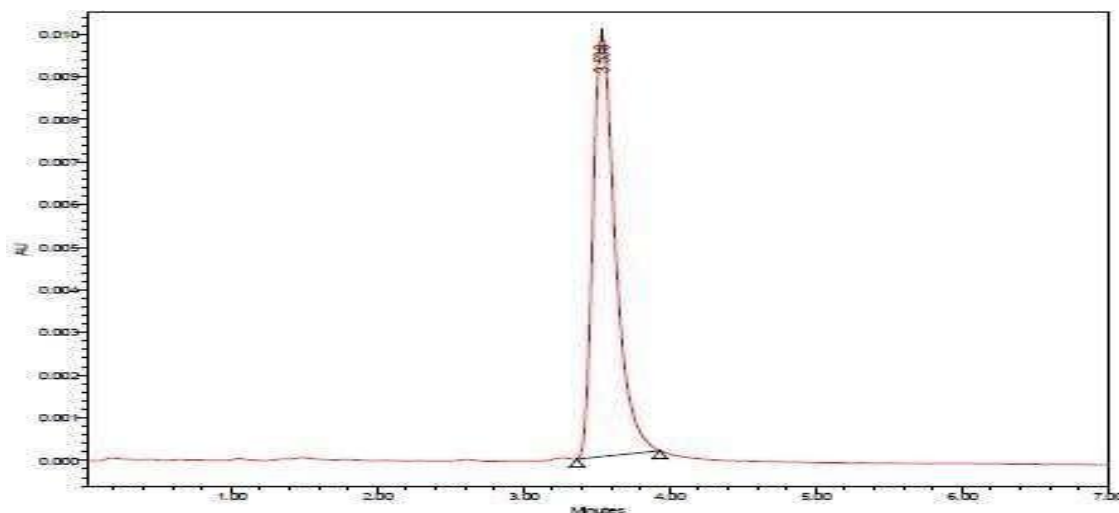
**Table No.9: linearity results**



**Fig-11: Chromatogram for Linearity-1 (6 ppm)**

**Table-32: Results of Linearity-1 (6 ppm)**

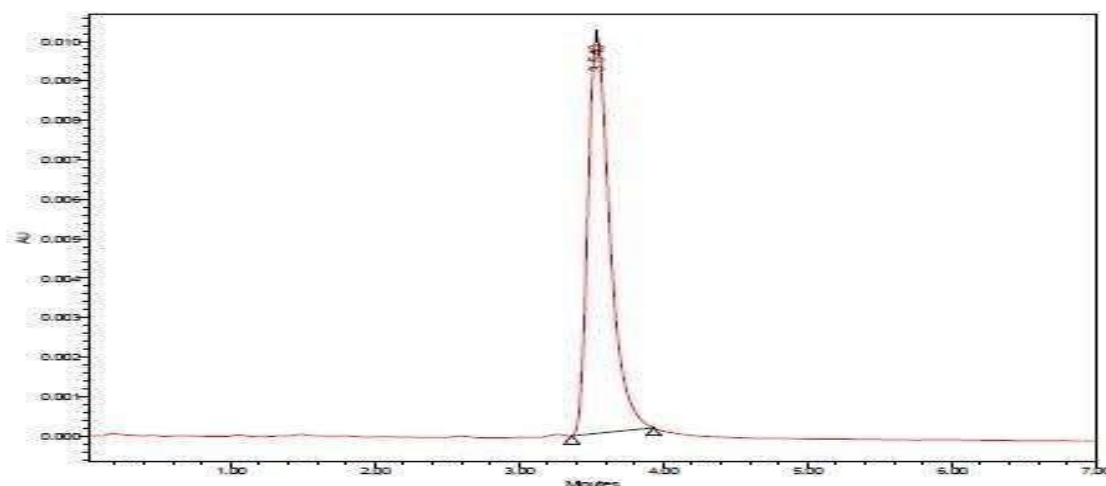
S.No.	Rt	Peak Area	Theoretical Plates	Tailing factor
1	3.543	370200	6854	1.17



**Fig-32: Chromatogram for Linearity-2 (8 ppm)**

**Table-33: Results of Linearity-2 (8 ppm)**

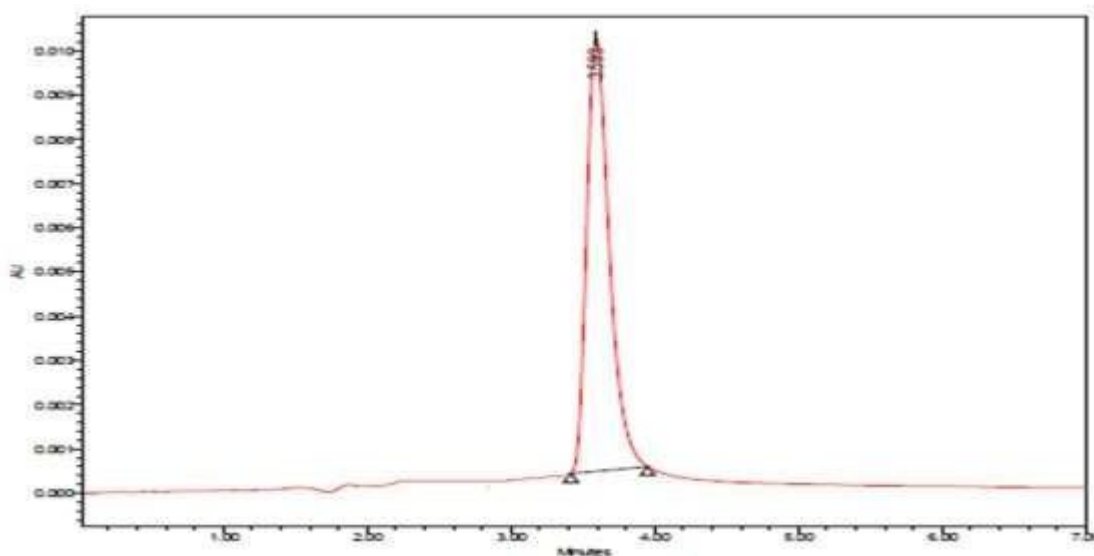
S.No.	Rt	Peak Area	Theoretical Plates	Tailing factor
1	3.538	490231	7458	1.21



**Fig-33: Chromatogram for Linearity-3 (10 ppm)**

**Table-34: Results of Linearity-3 (10 ppm)**

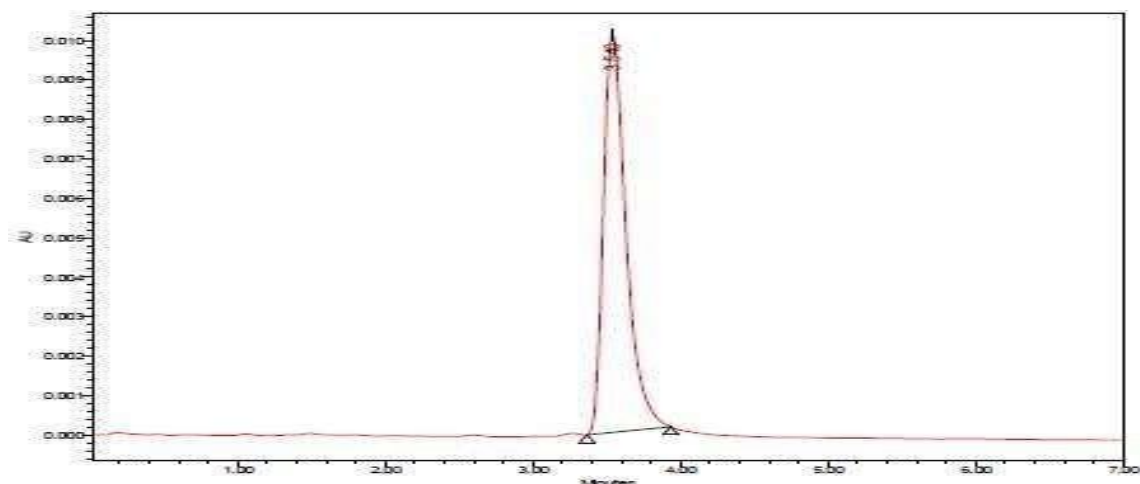
S.No.	Rt	Peak Area	Theoretical Plates	Tailing factor
1	3.543	602707	4286	1.20



**Fig-34: Chromatogram for Linearity-4 (12 ppm)**

**Table-35: Results of Linearity-4 (12 ppm)**

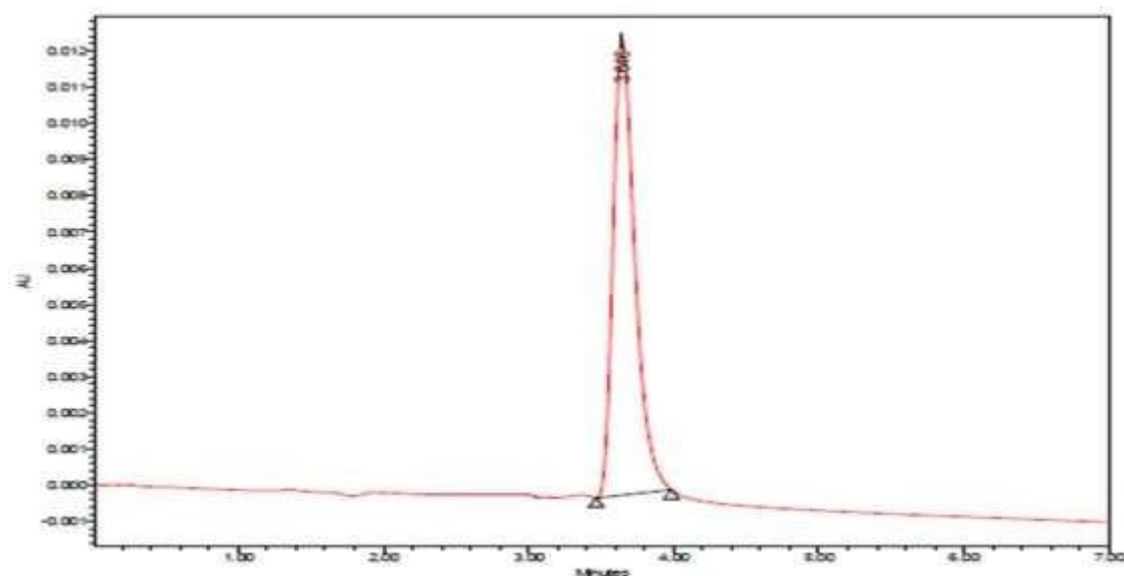
S.No.	Rt	Peak Area	Theoretical Plates	Tailing factor
1	3.593	717538	5896	1.26



**Fig-35: Chromatogram for Linearity-5 (14 ppm)**

Table-36: Results of Linearity-5 (14 ppm)

S.No.	Rt	Peak Area	Theoretical Plates	Tailing factor
1	3.543	829248	5486	1.42



**Fig-36: Chromatogram for Linearity-6 (16 ppm)**

Table-37: Results of Linearity-6 (16 ppm)

S.No.	Rt	Peak Area(	Theoretical Plates	Tailing factor
1	3.646	947852	4265	1.12



## **ROBUSTNESS**

Influence of small changes in chromatographic conditions such as flow rate, temperature, wavelength of detection & Acetonitrile content in mobile phase studied to determine the robustness of the method are also in favour of the developed RP-HPLC method for the analysis of fluconazole (API).

Change in parameter	% RSD
Flow (1.1 ml/min)	0.52
Flow (0.9 ml/min)	0.56
Temperature (27°C)	0.52
Temperature (23°C)	0.49
Wavelength of Detection (270 nm)	0.97
Wavelength of detection (266 nm)	0.98

Table No.10: results of method robustness

## **LOD & LOQ**

The minimum concentration level at which the analyte can be reliably detected (LOD) & quantified (LOQ) were found to be 0.90 & 2.90 µg/ml respectively.

## **RESULTS AND DISCUSSION**

### **Summary**

The optimum chromatographic conditions obtained from experiments can be summarised as below:

Mobile phase	ACN : Phosphate buffer (pH=3.2) =20:80
Column	Symmetry ODS RP C <sub>18</sub> , 5 µm, 15mm x 4.6mm i.d.

Flow rate	1.0 ml/ min.
Wavelength	274nm
Sampling System	Automatic
Temp. of Auto sampler	Ambient
Volume of injection	10µl
Run time	07 mins
Mode of Separation	Isocratic

Table No.11: summary of optimised chromatographic conditions

## Conclusion

A sensitive and selective RP-HPLC method has been developed and validated for the determination of fluconazole in pure form and pharmaceutical dosage form. The UV detection was performed at 274 nm. The retention time for fluconazole was found to be 3.544 minutes. The percentage standard deviation(%RSD) of the fluconazole was found to be 1.63%. the detector response was linear in the concentration range of 0-16µg/ml. The respective linear regression equation being  $Y=58945.x + 9634$  with  $R^2=0.999$ . % recovery values was found to be within range. Limit detection and limit quantitation was found to be 0.90µg/ml and 2.90µg/ml respectively. Further the process was rapid, accurate, precise. The rapid method was developed and validated according to the international council of harmonization (ICH) guidelines. The results of the study of fluconazole was found to be precise, accurate, simple, rapid which can be used for the routine determination of fluconazole in pure form and pharmaceutical dosage form.

**REFERENCE:**

1. Cuatrecasas P, Wilchek M, Anfinsen CB. Selective enzyme purification by affinity chromatography. *Proc Natl Acad Sci U S A*. 1968; 61:636–43.
2. Porath J. From gel filtration to adsorptive size exclusion. *J Protein Chem*. 1997; 16:463–8.
3. Harris DC. Exploring chemical analysis. 3rd ed. WH. Freeman &Co; 2004.
4. Gerberding SJ, Byers CH. Preparative ion-exchange chromatography of proteins from dairy whey. *J Chromatogr A*. 1998; 808:141–51.
5. Donald PL, Lampman GM, Kritz GS, Engel RG. Introduction to organic laboratory techniques. 4th ed. Thomson Brooks/Cole; 2006. pp. 797–817.
6. Harwood LM, Moody CJ. Experimental organic chemistry: Principles and Practice. Oxford: Blacwell Science; 1989:180–5.
7. Das M, Dasgupta D. Pseudo-affinity column chromatography based rapid purification procedure for T7 RNA polymerase. *Prep Biochem Biotechnol*. 1998; 28:339–48.
8. Karlsson E, Ryden L, Brewer J Protein purification. Principles, High Resolution Methods, and Applications. Ion exchange chromatography. 2nd ed. New York: Wiley; 1998.
9. Amercham Biosciences. Ion Exchange chromatography, Principles and methods,  
Amercham Pharmacia. Biotech SE. 2002;751
10. Walls D, Loughran ST. Protein chromatography: Methods and protocols, methods in molecular biology. 2011;681

11. Helmut D. Gel Chromatography, gel filtration, gel permeation, molecular sieves: a laboratory hand book. Springer-Verlag; 1969.
12. Mahn A, Asenjo JA. Prediction of protein retention in hydrophobic interaction Chromatography. Biotechnol Adv. 2005; 2:359–68.
13. Queiroz JA, Tomaz CT, Cabral JM. Hydrophobic interaction chromatography proteins. J Biotechnol. 2001; 87:143–59.
14. Porath J. Immobilized metal ion affinity chromatography. Protein ExpPurif. 1992; 3:263–81
15. Regnier FE. High-performance liquid chromatography of biopolymers. Science. 1983:245–52.